

Biotechnological and Commercial Utilization of Algae: A Review

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Abstract:

Algae are a diverse group of photosynthetic organisms found in marine and freshwater environments. With a rapid doubling time, they rank among the fastest-growing organisms. Algae utilize various pathways to capture atmospheric carbon dioxide and efficiently convert nutrients into biomass. Recently, their potential for food and fuel production has garnered significant attention. In the energy sector, algae-based biofuels have emerged as sustainable, eco-friendly, and cost-effective alternatives to traditional fuels, including bioethanol, biogas, biohydrogen, biodiesel, and bio-oil. Additionally, algae have been explored for food-related applications, such as producing single-cell proteins, pigments, bioactive compounds, pharmaceuticals, and cosmetics. This review underscores the extensive potential of algae for food and fuel industries while providing insights into commercially available algae-based products.

Keywords: Algae, Microalgae, Macroalgae, Biofuel, commercial, Potential

Introduction:

Algae have long been utilized for their ability to produce high biomass in extreme environments, outperforming cereal-based crops. Classified as third-generation biofuels, algae offer numerous advantages over agricultural crops for energy production. While the concept of using algae for biofuel is not new, the high production costs remain a challenge due to limited cultivation systems. However, advancements in technology are enabling largescale algae cultivation year-round across diverse climatic zones, from tropical to moderate regions.

Algae also have the unique ability to sequester carbon-rich gases, acting as effective absorbers of carbon dioxide and nitrogen oxides from various sources. Remarkably, 1 kg of algae biomass can fix approximately 1.8 kg of carbon dioxide. Furthermore, algae can grow in wastewater rich in nitrogen and phosphorus, simultaneously producing biofuels and reducing excess nutrient levels in the water[4,5,6].

A vast array of metabolites derived from algae, rich in bioactive compounds, remains largely untapped. For instance, *Haematococcus pluvialis*, a freshwater alga, is a key source of the commercially valuable pigment astaxanthin, while *Chlorella vulgaris* is widely used as a food supplement, and *Dunaliella* species are known for β -carotene production. Marine algae biomass can serve as a versatile feedstock for producing various fuels, including bioelectricity through co-firing, bioethanol, biodiesel, bio-oil via pyrolysis, and biomethane through fermentation.



The market potential for algal biofuels is immense, thanks to their sustainable approach to replacing fossil fuels. Algae typically contain over 50% starch, which can be converted into ethanol. Their cell walls, rich in carbohydrates, make them a viable raw material similar to cellulosic ethanol. Through pyrolysis, algal biomass can also be transformed into organic liquids, such as acetic acid, acetone, and methanol, along with clean and cost-effective gaseous products. Key factors that make algae a strong candidate for bioenergy production include high biomass productivity, ease of mechanical harvesting, and cost-effective production compared to other biomass sources.

Various Algae-Based Fuels:

In the modern age, the depletion of fossil fuels and the challenges of global warming have shifted global focus toward generating bioenergy from algal biomass. Enhancing energy access and ensuring energy security are now pivotal strategies for alleviating poverty. Currently, producing biofuels from algal biomass stands as the most viable alternative to reduce fossil fuel dependence and consumption. (Figure 1)

Biofuels

Biofuels refer to solid, liquid, or gaseous fuels derived from renewable raw materials. The effectiveness of any conversion process depends on factors such as the type and quantity of biomass, the desired energy output, and the economic value of the final product[9]. Agricultural crops are categorized as first-generation biofuels because they are used for food or feed; however, this creates a competition between food and fuel production, limiting the capacity of these biofuels to significantly contribute to overall fuel consumption.

Unlike biofuels from agricultural feedstocks, algae cultivation does not require agricultural land, avoiding competition with food production. Algae offer several advantages, including high biomass productivity, the ability to treat wastewater, year-round production, and adaptable chemical composition. Furthermore, the oil content of algae can be optimized by modifying cultivation techniques, enhancing their efficiency as a biofuel source.

Bio-Oil

Bio-oil is produced through thermo-chemical conversion, a process that transforms biomass into oil, along with char and gas, in the absence of oxygen at high temperatures. Due to its similarity to petroleum oils, bio-oil can serve as a viable substitute[11]. This conversion process involves two main steps: pyrolysis and thermo-chemical liquefaction[12].

Pyrolysis occurs at very high temperatures (350–530°C) and produces three fractions: liquid, gas, and solid. The liquid fraction consists of aqueous and non-aqueous phases, collectively referred to as bio-oil or tar, and requires pre-dried biomass. In contrast, thermo-chemical liquefaction processes wet biomass at lower temperatures (around 300°C) and high pressure (approximately 10 MPa).

Bio-oil contains various organic compounds derived from lipids, proteins, and carbohydrates in algae. Compared to lipids alone, algae yield a higher overall bio-oil output. Studies have explored the use of microalgae for bio-oil production through pyrolysis or thermal liquefaction. For instance, pyrolysis of *Dunaliella* biomass produces hydrocarbons, while bio-oil yields from various algae species include:

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- Up to 41% for *Spirulina*[18]
- 24%–45% for *Scenedesmus*[19]
- Approximately 37% for *Dunaliella*[20]
- Up to 49% for *Desmodesmus*[21]

Macroalgae also exhibit significant potential. Liquefaction of macroalgal biomass can yield up to 23% bio-oil[22, 23], with energy recovery reaching 63% in *Laminaria saccharina*[24] and bio-oil yields as high as 79% after hydrothermal liquefaction[25]. Conversely, freshwater macroalgae, such as *Oedogonium* and *Cladophora*, produce lower yields of 26% and 20%, respectively[26].

Biodiesel

In recent years, biodiesel has gained widespread attention and is primarily produced from oilseed crops such as soybean, palm, and rapeseed oils[27]. The cost of biodiesel production largely depends on the choice of raw material, which accounts for 50–85% of the total fuel price. For cost-effective production, it is crucial to evaluate feedstock based on productivity, quality, and the potential utilization of by-products[28, 29].

The process of converting lipids, mainly triacylglycerols or free fatty acids, into non-toxic and eco-friendly biodiesel is known as transesterification. In this process, crude algal oil, which has high viscosity, is transformed into low molecular weight fatty acid alkyl esters. Using a catalyst, the crude oil reacts with an alcohol, typically methanol, to produce fatty acid methyl esters (FAME) and glycerol as by-products. While acid catalysts have certain advantages[30], alkali catalysts are widely used due to their efficiency, being up to 400 times faster[31].

Species like *Chlorella vulgaris* and *Chlorella protothecoides*, known for their high oil content, have been extensively studied for biodiesel production. Gülyurt et al.[32]investigated the potential of *Chlorella protothecoides* using microwave-assisted transesterification. Optimum conditions for maximum biodiesel yield included a 9:1 methanol-to-oil molar ratio, 1.5% potassium hydroxide catalyst-to-oil ratio, and a reaction time of 10 minutes.

Microalgal biodiesel primarily consists of unsaturated fatty acids[33]. Algal biomass obtained from wastewater contains a mixture of algae species, resulting in varied fatty acid profiles. For example, Bjerk[43]successfully produced biodiesel using a blend of algae species, including *Chlorella sp.*, *Euglena sp.*, *Spirogyra sp.*, *Scenedesmus sp.*, *Desmodesmus sp.*, *Pseudokirchneriella sp.*, *Phormidium sp.*, and *Nitzschia sp.*.

Bioethanol

The production of bioethanol from algae has gained significant attention due to their high biomass productivity, diversity, variable chemical composition, and efficient photosynthetic rates. Algae are considered an ideal source for bioethanol production due to their high carbohydrate and polysaccharide content, as well as their thin cellulose walls. Two primary methods are employed for bioethanol production from algae: fermentation and gasification[42].

In many countries, commercial ethanol is produced on a large scale from sugary and starchy crops through fermentation. For algal bioethanol, the biomass is ground, and starch is



converted to sugars through enzymatic, acidic, or alkaline methods. Yeast, typically *Saccharomyces cerevisiae*, is added to initiate fermentation, converting sugars into ethanol[43]. The ethanol produced is then extracted and transferred to a distillation unit for further purification.

Research highlights several promising approaches:

- Ueno et al.[44]examined the marine green alga *Chlorococcum littorale* for ethanol production via dark fermentation, utilizing 27% of cellular starch within 24 hours at 25°C.
- John et al.[45]evaluated algal biomass as a renewable feedstock for bioethanol, emphasizing its potential as a sustainable biofuel. Species such as *Dunaliella*, *Chlorella*, *Chlamydomonas*, *Arthrospira*, *Sargassum*, *Spirulina*, *Gracilaria*, *Prymnesium parvum*, *Euglena gracilis*, and *Scenedesmus* have been explored.
- El-Sayed et al.[46]investigated bioethanol production from the seaweed *Ulva lactuca* using fermentation with yeast. Optimization through the Plackett-Burman design and immobilization techniques showed a conversion efficiency of 47.1% for free yeast and 52% for immobilized yeast.

Additionally, Obata et al.[47]studied bioethanol production from brown seaweeds (*Ascophyllum nodosum* and *Laminaria digitata*). These were pretreated with dilute sulfuric acid and hydrolyzed using commercial enzymes, yielding fermentable sugars such as glucose and rhamnose. Fermentation with non-conventional yeast strains, *Scheffersomyces stipitis* and *Kluyveromyces marxianus*, produced ethanol yields of approximately 6 g/L, with *K. marxianus* yielding slightly better results than *S. stipitis*.

Biobutanol Production

In regions such as Asia, Europe, and South America, algae cultivation is primarily focused on producing bioethanol and biogas. However, in the United States, algae are gaining recognition for their potential in biobutanol production. For over a century, butanol has been utilized as a transportation fuel and is now considered a promising biofuel. It is seen as a superior alternative to ethanol as a petroleum additive due to its low vapor pressure and high energy density[59].

The production of butanol has economic advantages, as the bacteria involved can digest not only starch and sugars but also cellulose found in algal biomass, making it comparable in cost to ethanol production[60]. Anaerobic fermentation by various *Clostridium* species can produce acetone, butanol, and ethanol (ABE) through a process known as ABE fermentation, which utilizes both hexose and pentose sugars[61]. However, butanol production faces challenges due to fermentation inhibition, which limits yield and productivity.

For instance, butanol has been produced from the fermentation of the alga *Ulva lactuca* using *Clostridium* strains, though yields are relatively low at 0.16 g of butanol per gram of algae, compared to ethanol under similar conditions[49]. Pretreatment of *Ulva lactuca* with hot water, followed by enzymatic hydrolysis using commercial cellulases, has shown promise in producing ABE compounds, achieving a yield of 0.35 g of ABE per gram of sugar[61].

Algae-Based Non-Energy Applications



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Algae have immense potential for a wide range of products due to their vast diversity and the ability to modify their chemical composition based on cultivation conditions. However, the exploration of algae-based products remains limited, leaving much of this natural resource untapped. Several factors contribute to this limitation, including economic constraints, regulatory hurdles for product approvals[63], limited public awareness of algae-based products[62], insufficient knowledge of algae cultivation techniques, and inadequate investments in large-scale production facilities.

Among the commercially available algae-derived products, most are used in food applications or as alginates, which are primarily sourced from seaweed. These are moderately derived from natural populations rather than cultivated algae (Table 1).

Pharmaceuticals

Algae are abundant in unique biologically active compounds, including both primary and secondary metabolites, making them a promising resource for the pharmaceutical industry[65]. These bioactive compounds are likely a result of the algae's natural habitat in aquatic ecosystems, where they face competitive pressures between producers and consumers. Microalgae, in particular, are a rich source of bioactive compounds that can be harnessed for commercial use. Algae provide a diverse array of pharmaceutical products, proteins, vaccines, and nutrients that are otherwise difficult or costly to produce from animal or plant sources[66, 67]. While the commercialization of pharmaceutical products derived from microalgae is still in its early stages, it holds great potential to become a multibillion-dollar industry in the near future. With their remarkable genetic potential to produce various bioactive compounds, microalgae are positioned at the forefront of biotechnological research.

Ĺ	Table 1. Useful substances present in		
S.No.	Natural substances present inalgae	Different compounds derived from algae	
1.	Pigments	Astaxanthin, lutein, zeaxanthin, canthaxanthin, chlorophyll, phycocyanin, phycoerythrin, fucoxanthin	
2.	Carotenoids	β-carotene	
3.	Polyunsaturated fatty acids (PUFAs)	DHA, EPA, ARA, GAL	
4.	Vitamins	Biotin, riboflavin, nicotinic acid, pantothenate, folic acid	
5.	Antioxidants	Catalases, polyphenols, superoxide dismutase, tocopherols	
6.	Other	Antimicrobial compounds, toxic products, aminoacids, proteins	

Various unicellular algae, including *Chlorella vulgaris* and *Chlamydomonas pyrenoidosa*, have demonstrated antibacterial properties against a range of pathogens, including both grampositive and gram-negative bacteria, through their cell extracts and growth media extracts. Additionally, there have been some reports of in vitro antifungal activity from the extracts of green algae and diatoms. Certain blue-green algae and microalgae, such as *Ochromonas sp.*

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and *Prymnesium parvum*, are known to produce toxic substances that hold significant potential for pharmaceutical applications[70, 71]. Many species of *Cyanobacteria* are recognized for producing intracellular and extracellular metabolites with a broad spectrum of biological activities, including antibacterial, antifungal, and antiviral effects[72]. (Table 2)

Table 2: Biotechnological applications of bioactive

S.No.	Algae Specis	Different compounds	Uses
1.	Spirulina platensis	Phycocyanins	Nutraceuticals, cosmetics
2.	Chlorella vulgaris	Ascorbic acid	Health food, food supplement, food surrogate
3.	Haematococcus pluvialis	Carotenoids, astaxanthin	Nutraceuticals,, pharmaceuticals, additives
4.	Odontella aurita	Fatty acids	Pharmaceuticals, cosmetics, baby food
5.	Porphyridium cruentum	Polysaccharides	Pharmaceuticals, cosmetics,
6.	Dunaliella salina	Carotenoids	Nutraceuticals,, food supplement, feed

Cosmetics

Microalgae species, such as *Arthrospira* and *Chlorella*, are well-established in the skincare market, with companies like LVMH (Paris, France) and Daniel Jouvance (Carnac, France) developing their own microalgal cultivation systems. Microalgal extracts are included in various cosmetic products, including anti-aging creams, rejuvenating treatments, sun protection products, and hair care items[86]. As awareness of skin cancer and photoaging caused by sun exposure has increased, there has been a growing demand for sunscreen products in recent years.

Amino acids, such as mycosporine, have gained commercial interest for their natural UV-blocking properties in sunscreens. Many microalgae produce metabolites like sporopollenin, scytonemin, and mycosporine, which protect them from UV radiation while allowing visible light, essential for photosynthesis, to pass through[87]. Algae components are also commonly used as thickening agents, water-binding agents, and antioxidants in cosmetics[88]. Various algae extracts are found in face and skin care products, with species such as *Chondrus crispus*, *Ascophyllum nodosum*, *Alaria esculenta*, *Spirulina platensis*, *Nannochloropsis oculata*, *Chlorella vulgaris*, and *Dunaliella salina* being utilized.

For example, *Spirulina* is featured in beauty products like the *Spirulina Firming Algae Mask* by Optimum Derma Aciditate, which improves moisture balance and enhances skin immunity, and the *Spirulina Whitening Facial Mask* by Ferenes Cosmetics, which contains proteins and herbal extracts to improve skin complexion and reduce wrinkles without side effects. Codif Recherche & Nature (Paris, France) markets a product made from *Phormidium persicinum*, called *Phormiskin Bioprotech G*, known for its unique photo-protective properties[89]. (Table 3)

Pigments

Microalgae are rich in pigments associated with light absorption, including chlorophyll (the primary photosynthetic pigment), phycobiliproteins, and carotenoids. Carotenoids extracted from microalgae are widely used in the market. For instance, β -carotene from Dunaliella serves as a vitamin supplement in health foods,



while *lutein*, *zeaxanthin*, and *canthaxanthin* are used in pharmaceuticals and for enhancing chicken skin color. *Astaxanthin*, extracted from *Haematococcus*, is used in aquaculture to provide a natural red color for fish like salmon.

Phycobiliproteins, such as phycocyanin and phycoerythrin, which are unique to algae, are already utilized in food and cosmetics[90, 91]. The antioxidant properties of carotenoids are particularly important for human use, as they act as free radical scavengers and have anti-cancer effects. Among natural antioxidants, *astaxanthin* stands out for its potent effects. Due to its antioxidant properties and role as a precursor to vitamin A, β -*carotene* is now commonly used in health foods. Pigments extracted from microalgae are also commercially used as natural food colorants.

Table 3: Sources of UV-screening compounds from

S.No.	UV screening compound	Algae	
1.	Sporopollenin	Characium terrestre, Coelastrum microporum, Enallax coelastroides, Scenedesmus sp., Scotiellopsis rubescens, Dunaliellasalina, Chlorella fusca	
2.	Scytonemin	Chlorogloeopsis sp., Calothrix sp., Scytonema sp., Nostoccommune, Nostoc punctiforme	
3.		Ankistrodesmus spiralis, Chlorella minutissima, Chlorella sorokiniana, Dunaliella tertiolecta, Isochrysis sp., Corethron cri- ophilum, Stellarima microtrias, Alexandrium catenella	



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colorants and cosmetic ingredients.

β-Carotene

Carotenoids are commonly used as natural food colorants, additives in cattle feed, and in various cosmetics. Nutritionally, carotenoids like β -carotene serve as a provitamin A[68]. The physical properties of natural β -carotene make it a preferable choice over synthetic alternatives. Specifically, β -carotene is fat-soluble. Recently, the National Cancer Institute recognized β -carotene as an anticancer agent, and numerous studies have highlighted its effectiveness in managing cholesterol and reducing the risk of heart disease. These new findings position β -carotene as a promising product with potential for increased demand in the commercial market.

Astaxanthin

Astaxanthin is another carotenoid derived from microalgae, with various industrial applications. This pigment, a keto-carotenoid, is primarily obtained from the alga *Haematococcus pluvialis* and is produced under stress conditions. During these conditions, the algae cells transition from thin, flagellated forms to red, thick-walled resting stages, making up about 4-5% of the dry weight[96]. Astaxanthin is commonly used as a food colorant, a feed additive in the poultry industry, and as a supplement for fish such as salmon, trout, and shrimp[96].

Phycobiliproteins

The levels of various elements, such as phycobiliproteins, in algae are influenced by environmental factors like light intensity and the spectral quality of light. For instance, when *Spirulina platensis* is cultivated under different light intensities, the amount of phycocyanin ranges from 11% to 12.7% of dry weight[101]. *Spirulina* and *Porphyridium* are two common microalgae commercially cultivated for phycobiliprotein production. These pigments have significant potential in the food industry as natural colorants, in cosmetics, and as diagnostic tools in biomedical research, such as fluorescent markers[102]. For example, phycocyanin is marketed under the brand name "Lina Blue" by Dainippon Ink and Chemicals for use in products like popsicles, candies, cold drinks, dairy items, and chewing gum. The price of phycobiliproteins can range from US\$ 3 to US\$ 1500 per mg for certain cross-linked pigments[103].

Algae-derived compounds for human usage

The use of algae as a food has been dated back to 2500 year ago in Chinese literature [108]. Many macroalgae (seaweeds) are consumed in various parts of Asia as food directly and edible in small amount by native people of countries like Africa, South America and Mexico due to their vitamins and nutritional value [109]. They can also be added into different foods like pastas, snacks, gums, beverages, noodles, cookies [110, 111]. A blue green alga named Spirulina platensis is acquiring worldwide attention as food additive due to its high nutritional value as a food to human. It has been demonstrated as a rich source of proteins [112], polyunsaturated fatty acids [113], pigments [114, 115], vitamins and phenolics [112, 116]. These days Chlorella is also being majorly sold out in food store and as a fish feed, like Spirulina [117]. Currently, the microalgal market is ruled by Chlorella and Spirulina [118, 68], due to their high protein value, nutritional value and ease of growing. Their biomass is sold as tablets, capsules and liquids, to be used as nutritional supplement [118]. The biomass of microalgae composed of three major portions: proteins, carbohydrates, and lipids [119]. In the table 5, compositional analysis of various microalgal biomass in terms of main components is given.



S.No.	Product group	s high-value compour 	Examples (producer)
1.	Phycobiliproteins	Pigments, cosmetics, vitamins	Phycocyanin (Spirulina platensis)βcaroteneαstaxanthin(Haematococcus pluvialis)
2.	Polyunsaturated fatty acids (PUFAs)	Nutraceuticals, foo dsupplements	EPA-(Chlorella minutissima) DHA- (Schizochytrium sp.)
3.	Vitamins	Nutrition	Biotin -(Euglena gracilis) α-tocopherol) -(Euglena gracilisa Vitamin C- (Prototheca moriformis)

Table 5. Commercialization of important microalgae for nutrition (Adapted from

S.No.	Algae	Algae based companies	Different algae products
1.	Spirulina (Arthrosphira)	Hainan Simai Pharmacy Co. (China) Earthrise Nutritionals (California, USA) Cyanotech Corp. (Hawaii, USA)	powders, tablets, powders, tablets, powders, beverages, tablets, chips, pasta
2.	Chlorella	Taiwan Chlorella Manufacturing Co.(Taiwan)	tablets, powders, nectar, noodles,powders
3.	Dunaliella salina	Cognis Nutrition and Health(Australia)	powders b-carotene

Fertilizer

Both macroalgae and microalgae contain various compounds that support plant growth, including promoting germination, leaf or stem development, and flowering, and can also serve as biological protectants against plant diseases[68]. These algae are used in several coastal regions. After extracting oil and carbohydrates from algae, a significant amount of nutrients remain in the spent biomass. This leftover biomass can be repurposed as a biofertilizer, enhancing the economic value of algae by allowing its reuse in cultivation after nutrient extraction. The remaining biomass can be utilized as fertilizer. Many cyanobacteria are capable of fixing atmospheric nitrogen, making them effective biofertilizers. They play a crucial role in preserving soil fertility and enhancing rice growth and yield when used as natural biofertilizers[121]. Nitrogen, after water, is the second most important factor for plant growth in fields, with fertilizers typically meeting this requirement[122]. Blue-green algae (BGA) such as *Nostoc*, *Anabaena*, and *Tolypothrix* are capable of fixing atmospheric nitrogen and are used as inoculants for growing paddy crops in both upland and lowland conditions[123].



Fibres for Paper

Sulfur-containing polysaccharides provide structural stability to most algae[124]. Algae that contain cellulose can potentially be used as a feedstock for paper production, although there are few examples of algae being utilized as a non-wood fiber source. For instance, algae collected from a municipal wastewater treatment plant were incorporated into a 10% pulp mix by Ververis et al.[125], which led to a significant increase in the mechanical strength of the paper and a 45% reduction in material costs due to decreased brightness. This resulted in a 0.9-4.5% reduction in overall costs. However, this concept remains largely within the research phase, and commercialization efforts need further attention.

Processed Food Ingredients

Agar, alginates, and carrageenans are some of the most valuable products derived from algae, known for their gelling and thickening properties. In recent years, there has been significant growth in algae research and development, particularly in areas such as protoplast fusion, macroalgal cell cultures, and transgenic algae[68].

Agar

Agar, obtained from macroalgae, has numerous applications in food products such as frozen foods, desserts, candies, and fruit juices. It is also used in various industries, including paper sizing, textile printing, and molecular biology as agarose. Additionally, it plays a role in biomedical fields, where it is used to produce capsules, tablets, and anticoagulants[125].

Carragenans

Carrageenan, a water-soluble polysaccharide derived from algae, is more commonly used than agar as an emulsifying and stabilizing agent in a variety of food products. κ - and ι -carrageenans are often utilized in items such as jellies, jams, desserts, and meat products due to their thickening properties. In addition, carrageenans have been explored for various pharmaceutical applications, including antiviral, antitumor, and anticoagulant effects.

S.No.	Cyanobacteria	Growth enhancers
1.	Cylindrospermum sp.	Vitamin B12
2.	Tolypothrix tenuis	Vitamin B12
3.	Nostoc muscorum, Hapalosiphon fontinalis	Vitamin B12
4.	Nostoc, Hapalosiphon	Auxin like Indole-3-acetic acid indole-3-propionic acid or 3-methyl indole

Alginate

Alginate, a compound derived from brown algae, is valued in the textile industry for sizing cotton yarn and is highly regarded for its gelling properties. Its chelating ability and capacity to form highly viscous solutions make it a promising candidate for use in the food and pharmaceutical industries.



Conclusion

In the field of algae, there is a pressing need to advance research and development efforts to overcome technological challenges, as algae have the potential to yield novel chemicals and bioactive compounds. The applications of algae are vast within the biotechnology sector, and it is essential to tap into their rich diversity for a wide range of uses. Algae show great promise as sources of biofuels, high-value molecules, nutraceuticals, and bioactive metabolites, which could lead to the discovery of new drugs. To fully realize their potential, there is a need for complete utilization of algal biomass and further exploration of their diverse applications.

References

- 1. Ackman, R. G., Tocher, C. S., McLachlan, J.(1968). Marine phytoplankton fatty acid. J Fisheries Res. Board of Canada. 25, 1603-1620
- 2. Adey, W. H., Loveland, K. (2007). Dynamic aquaria: building living ecosystems. 3rd Edn, Academic Press, New York, USA
- 3. Antoni, D., Zverlov, V. V., Schwarz, H. (2007). Biofuels from Microbes. Appl. Microbiol. Biotechnol.77, 23-35
- 4. Bilanovic, D., Shelef, G., Sukenik, A. (1988). Flocculation of microalgae with cationic polymers: effects of medium salinity. Biomass 17, 65-76
- 5. Benemann, J. R., Oswald, W.J. (1996). Systems and Economic Analysis of Microalgae Ponds for Conversion of CO2 to Biomass. Pittsburgh Energy Technology Center, USA. pp. 260.
- 6. Bigogno, C., Khozin-Goldberg, I., Boussiba, S., Vonshak, A., Cohen, Z. (2002). Lipid and fatty acid composition of the green oleaginous alga, Parietochloris incisa, the richest plant source of arachidonic acid. Phytochem. 605, 497- 503.
- 7. Olson, G. J., Ingram, I. (1975). Effects of Temperature and Nutritional Changes on the Fatty Acids of Amenellum Ouadrupicatum L. Bacteriol124, 373
- 8. Mata, T. M., Martins, A. A. et al. (2010)."Microalgae for biodiesel production and other applications: A review." Renewable & Sustainable Energy Reviews 14, 217-232.
- 9. Wout, R., Greenwell, H., Davies, D. (2013). Theodorou, M. Methods of Ensiling Algae, Ensiled Algae and Uses of Ensiled Algae. WO2013045931- A1
- 10. Awasthi, M., Singh, R. K. (2011). Development of algae for the production of bioethanol, biomethane, biohydrogen and biodiesel. Int J CurrSci. 1, 14-23
- 11. Kishimoto, M., Okakura, T., Nagashima, H., Minowa, T., Yokoyama, S. et al.(1994). CO2 Fixation and Oil Production Using Microalgae. JFermentation Bioeng. 78, 479-482
- 12. Demirbas, A. (2000). Mechanisms of liquefaction and pyrolysis reactions of biomass. Energy Conversion and Management 41, 633-646
- 13. Dote, Y., Sawayama, S., Inoue, S., Minowa, T., Yokoyama, S. (1994). Recovery of Liquid Fuel from Hydrocarbon-Rich Microalgae by Thermochemical Liquefaction. Fuel 73, 1855-1857
- 14. Sawayama, S., Minowa, T., Yokoyama, S. Y. (1999). Possibility of renewable energy production and CO2 mitigation by thermochemical liquefaction fmicroalgae. Biomass & Bioenergy 17, 33-39
- 15. Peng, W. M., Wu, Q. Y., Tu, P. G. (2000). Effects of temperature and holding time on production of renewable fuels from pyrolysis of Chlorella protothecoides. J Appl. Phycol. 12, 147-152
- 16. Peng, W. M., Wu, Q. Y., Tu, P. G. (2001a).

Pyrolytic characteristics of heterotrophic Chlorella protothecoides for renewable bio-fuel production. J Appl. Phycol. 13, 5-12



- Peng, W. M., Wu, Q. Y., Tu, P. G., Zhao, N. M. (2001b). Pyrolytic characteristics of microalgae as renewable energy source determined by thermogravimetric analysis. Bioresource Technology 80: 1-7 Phys. Chem. 14, 3140–3147
- 18. Jena, U., Das, K.C. (2011). Comparative evaluation of thermochemical liquefaction and pyrolysis for bio-oil production from microalgae. Energy Fuels. 25, 5472–5482
- 19. Vardon, D.R., Sharma, B.K., Blazina, G.V., Rajagopalan, K., Strathmann, T.J. (2012). Thermochemical conversion of raw and defatted algal biomass via hydrothermal liquefaction and slow pyrolysis. Bioresour. Technol. 109, 178–187.
- 20. Minowa, T., Yokoya, S. Y., Kishimoto, M., Okakura, T. (1995). Oil production from algae cells of Dunaliella Tereiolata by direct thermochemical liquefaction. Fuel 74, 1731-1738.
- 21. Alba, L.G., Torri, C., Samori, C., Van der Spek, J., Fabbri, D. et al. (2012). Hydrothermal treatment ofmicroalgae: Evaluation of the process asconversion method in an algae biorefinery concept. Energy Fuels. 26, 642–657.
- 22. Zhou, D., Zhang, L., Zhang, S., Fu, H., Chen, J. (2010). Hydrothermal liquefaction of macroalgae Enteromorpha prolifera to bio-oil. Energy Fuels. 24, 4054–4061
- 23. Murphy, F., Devlin, G., Deverell, R., McDonnell, K. (2013). Biofuel production in ireland—An approachto 2020 targets with a focus on algal biomass. Energies . 6, 6391–6412
- 24. Anastasakis, K., Ross, A.B. (2011). Hydrothermal liquefaction of the brown macro-alga laminaria saccharina: Effect of reaction conditions on product distribution and composition. Bioresour.Technol. 102, 4876–4883.
- 25. Bach, Q.V. ,Sillero, M.V., Tran, K.Q., Skjermo, J. (2014). Fast hydrothermal liquefaction of a norwegian macro-alga: Screening tests. Algal Res. doi:10.1016/j.algal.2014.05.009
- 26. Neveux, N., Yuen, A.K.L., Jazrawi, C., Magnusson, M., Haynes, B.S. et al. (2014). Biocrude yield and productivity from the hydrothermal liquefaction of marine and freshwater green macroalgae. Bioresour. Technol. 155, 334–341
- 27. Argonne National laboratory. (2008). Life-Cycle Assessment of Energy and Greenhouse Gas Effects of Soybean-Derived Biodiesel and Renewable Fuels
- 28. Nikolaison, L., Dahl, J., Bech, K.S., Bruhn, A., Rasmussen, M.B., et al. (2012). Energy production fom macroalgae. In Proceedings of the 20th European Biomass Conference, Milan, Italy, 18–22 June 2012.
- 29. Adams, J.M.M., Ross, A.B., Anastasakis, K., Hodgson, E.M., Gallagher, J.A. et al. (2011). Seasonal variation in the chemical composition of the bioenergy feedstock laminaria digitata for thermochemical conversion. Bioresour. Technol. 102, 226–234.
- 30. Meng, X., Yan, J., Xu, X., Zhang, L., Nie, Q. et al. (2009). Biodiesel production from oleaginous microorganisms. Renew Energy. 34, 1-5.
- 31. Miao, X. and Wu, Q. (2006). Biodiesel production from heterotrophic microalgal oil. Bioresour. Technol. 97, 841-846
- 32. Gülyurt, M.O., Özçimen, D., Benan, ¹I. (2016).Biodiesel Production from Chlorella protothecoides Oil by Microwave-Assisted Transesterification. Int.J. Mol. Sci.17, 579
- 33. Belosevic, S. (2010). Modeling approaches to predict biomass co-firing with pulverized coal. Open Thermodyn. J. 4, 50–70.
- 34. Bjerk, T. R. (2012). Microalgae cultive in fotobiorreator and joint reactor objectiving the bioremediation and production of biofuels Potential Production of Biofuel from Microalgae Biomass Produced in Wastewater http://dx.doi.org/10.5772/52439. 2012
- 35. Saifuddin, N., Priatharsini, P. (2016). Developments in Bio-hydrogen Production from Algae: A Review. Res. J. Appl. Sci. Eng.Technol 12,968-982



- 36. Prince, R. C., Kheshgi, H. S .(2005). The photobiological production of hydrogen: Potential efficiency and effectiveness as a renewable fuel. Critic. Rev. Microbiol. 31, 19-31
- 37. Hankamer, B., Lehr, F., Rupprecht, J., Mussgnug, J. H., Posten, C. (2007). "Photosynthetic biomass and H2 Production by Green Algae: From Bioengineering to Bioreactor Scale-Up," PhysiologiaPlantarum, 131,10-21
- 38. Guan, Q.Q. (2012).Savage, P.E.; Wei, C.H. Gasification of alga nannochloropsis sp in supercritical water. J. Supercrit. Fluids. 61, 139–145
- 39. Leu, S., Boussiba, S. (2014). Advances in the production of high-value products by microalgae. Industrial Biotechnology 10, 169-183
- 40. Chynoweth, D. P., Turick, C. E., Owens, J. M., Jerger, D. E., Peck, M. W. (1993). Biochemical methane potential of biomass and wastefeedstocks. Biomass and Bioenergy 5, 95-111
- 41. Wang, M., Lee, E., Dilbeck, M.P., Liebelt, M., Zhang, Q. et al. (2016). Thermal pretreatment of microalgae for biomethane production: experimental studies, kinetics and energy analysis. J Chem Tech. Biotechnol. 92, 399-407
- 42. Minowa, T. and Sawayama, S. (1999). A novelmicroalgal system for energy production with nitrogen cycling. Fuel 78, 1213-1215
- 43. Minowa, T., Yokoyama, S., Kishimoto, M., Okakura, T. (1995). Oil production from algal cells of Dunaliella-tertiolecta by direct thermochemical liquefaction. Fuel 74, 1735–1738
- 44. Ueno, Y., Kurano, N. and Miyachi, S. (1998). Ethanol production by dark fermentation in themarine green alga, Chlorococcum littorale. J Ferment Bioeng 86, 38-43.
- 45. John, R. P., Anisha, G., Nampoothiri, K.M. and Pandey, A. (2011). Micro and macroalgal biomass: a renewable source for bioethanol. Bioresour. Technol. 102, 186–193.
- 46. El-Sayed, W.M. M., Ibrahim, H.A.H., Abdul-Raouf, U.M., El-Nagar, M.M. (2016). Evaluation of Bioethanol Production from Ulva lactuca BySaccharomyces cerevisiae. J Biotechnol. Biomater.6, 226
- 47. Oluwatosin, O., Joseph, A., Heike, B., Graeme, W. (2016). Ethanol production from brown seaweed using non-conventional yeasts. Bioethanol 2, 134–145
- 48. Yoza, B.A. and Masutani, E.M. (2013). The analysis of macroalgae biomass found around Hawaii for bioethanol production. Environ. Technol.34, 1859–1867
- 49. Nikolaison, L., Dahl, J., Bech, K.S., Bruhn, A., Rasmussen, M.B. et al. (2012). Energy production fom macroalgae. In Proceedings of the 20th European Biomass Conference, Milan, Italy, 18–22 June 2012
- 50. Jung, K.A., Lim, S.R., Kim, Y., Park, J.M. (2013). Potentials of macroalgae as feedstocks for biorefinery. Bioresour. Technol., 135, 182–190.
- 51. Meinita, M.D.N., Marhaeni, B., Winanto, T., Jeong, G.T., Khan, M.N.A. et al. (2013). Comparison of agarophytes (Gelidium, Gracilaria, and Gracilariop- sis) as potential resources for bioethanol production. J. Appl. Phycol. 25, 1957–1961.
- Roesijadi, G., Copping, A.E., Huesemann, M.H., Foster, J., Benemann, J.R. (2010). Techno-Economic Feasibility Analysis of Offshore Seaweed Farming for Bioenergy and Biobased Products; PNNL-19944; U.S. Department of Energy: Washington, DC, USA, 2010.
- 53. Wargacki, A.J.; Leonard, E.; Win, M.N.; Regitsky, D.D.; Santos, C.N.S.; Kim, P.B.; Cooper, S.R.; Raisner, R.M.; Herman, A.; Sivitz, A.B.; et al. An engineered microbial platform for direct biofuel production from brown macroalgae. Science **2012**,335, 308–313
- 54. Aizawa, M., Asaoka, K., Atsumi, M., Sakou, T. (2007). Seaweed Bioethanol Production in Japan— The Ocean Sunrise Project. In Proceedings of the OCEANS 2007, Vancouver, BC, Canada, 29 September–4 October 2007; pp. 1–5.



- 55. Yoza, B.A., Masutani, E.M. (2013). The analysis of macroalgae biomass found around Hawaii for bioethanol production. Environ. Technol. 34, 1859–1867.
- 56. Horn, S.J. and Aasen, I.M. (2000). Ostgaard, K. Ethanol production from seaweed extract. J. Ind. Microbiol. Biotechnol. 25, 249–254
- 57. Walker, D.A. (2010). Biofuels—for better or worse?Ann. Appl. Biol. 156, 319–327
- 58. Balat, M., Balat, H. and Oz, C. (2008). Progress in bioethanol processing. Prog. Energy Combust. Sci. 34, 551–573
- 59. Potts, T., Du, J., Paul, M., May, P. and Beitle, R. (2012). Hestekin, J. The production of butanol from Jamaica bay macro algae. Environ. Prog.Sustain. Energy 31, 29–36
- 60. Gao, K., Orr, V., Rehmann, L. (2016). Butanol fermentation from microalgae-derived carbohy- drates after ionic liquid extraction. Bioresour. Technol. 206, 77–85
- 61. Wal Hetty V. D., Bram, L.H.M., Sperber, B. T., Robert, R.C., Bakker, W. B. et al. (2013).Production of acetone, butanol, and ethanol from biomass of the green seaweed Ulva lactuca. Bioresour. Technol. 128, 431-437
- 62. Edwards, M. (2008). Green Algae Strategy End Biowar I and Engineer Sustainable Food and Biofuels. Tempe, Arizona, USA, LuLu Press
- 63. Reith, J.H. (2004) Sustainable Co-Production of Fine Chemicals and Energy from Microalgae: Public Final E.E.T
- 64. Priyadarshani, I. and Biswajit, R. (2012). Commercial and industrial applications of microalgae A review J. Algal Biomass Utln. 3, 89–100
- 65. Rania, M.A. and Hala, M.T. (2008). Antibacterial and antifungal activity of Cynobacteria and green Microalgae evaluation of medium components by Plackett-Burman design for antimicrobial activity of Spirulina platensis. Global Journal of Biotechnol. Biochem. 3, 22-31.
- 66. Metting, B and Pyne, J.W. (1986) Biologically-Active Compounds from Microalgae. Enzyme and Microbial Technology 8, 386-394
- Borowitzka, M. A. (1988). Micro-algal biotechnolo- gy. Cambridge University Press, Cambridge, pp. 477
- 68. Bhattacharjee, M. (2016). Pharmaceutically valuable bioactive compounds of algae. Asian J Pharm Clin Res. 9, 43-47
- 69. Olaizola, M. (2003). Commercial development of microalgal biotechnology: from the test tube to the marketplace. Biomol. Eng. 20, 459-466
- 70. Borowitzka, M.A. and Borowitzka, L.J. (1992). In : Microalgal Biotechnology eds. Borowitzka M.A. pp. 27- 58. Cambridge, Cambridge University Press.
- 71. Katircioglu, H., Beyatli, Y., Aslim B., Yuksekdag, Z., Atici, T. (2006). Screening for antimicrobial agent production in fresh water. Internet J. Microbiol. 2, 187-197
- Noaman, N.H., Fattah, A., Khaleafa, M. and Zaky,
 S.H. (2004). Factors affecting antimicrobial activity of Synechococcus leopoliensis. Microbiolog- ical Research. 159, 395-402.
- 73. Priyadarshani, I. and Biswajit, R. (2012). Commercial and industrial applications of microalgae A review J. Algal Biomass Utln. 3, 89–100
- 74. Patterson, G.M.L., Baker, K.K., Baldwin, C.L., Bolis, C.M. and Caplan, F.R.. et al. (1993). Antiviral activity of cultured blue-green algae (Cyanophyta). J Phycol. 29, 125-130.
- 75. Dey, B., Lerner, D.L., Lusso, P., Boyd, M.R. and Elder, J.H. (20000). Multiple antiviral activities of cyanovirin-N: blocking of human immunodeficiency virus Type 1 gp120 interaction with CD4 and coreceptor and inhibition of diverse enveloped viruses. J Virol. 74, 4562-4569



- 76. Tsai, C.C., Emau, P., Jiang, Y., Agy, M.B., Shattock, R.J. et al. (2004). Cyanovirin-N inhibits AIDS virus infections in vaginal transmission models. AIDS Res Human Retrovir. 20, 11–18.
- 77. Hayashi, K., Hayashi, T. and Kojima, I. (1994). A natural sulfated polysaccharide, calcium spirulan, isolated from Spirulina platensis in vitro and exvivo evaluation of anti-herpes simplex virus and anti-human immunodeficiency virus activities. AIDSRes Hum Retrovir, 12, 1463-1471.
- 78. Ayehunie, S., Belay, A., Baba, T.W. and Ruprecht, R.M. (1998). Inhibition of HIV- 1 replication by an aqueous extract of Spirulina platensis. J Aquir Immun Defic Syndr Hum Retrovirol. 18, 7-12
- 79. Huleihel, M., Ishamu, V., Tal, J. and Arad, S.M. (2001). Antiviral effect of red microalgal polysaccharides on Herpes simplex and Varicella zoster viruses. J Appl Phycol. 13, 127-134.
- 80. Patterson, G.M.L., Baldwin, C.L., Bolis, C.M., Caplan, F.R. and Karuso, H. et al. (1991). Antineoplastic activity of cultured blue-green algae (Cyanobacteria). J Phycol 27, 530-536
- 81. Gerwick, W.H., Roberts, M.A., Proteau, P.J., Chen, J.L. (1994). Screening cultured marine microalgae for anticancer type activity. J Appl Phycol. 6, 143-149.
- 82. Boopathy, N.S. and Kathiresan, K.(2010). Anticancer drugs from marine flora: An overview. J.Oncol. 4, 1-18.
- Furusawa, E., Moore, R.E., Mynderse, J.S., Norton,
 T.R. and Patterson, G.M.L. (1994). New purified culture of Scytonema pseudohofmanni ATCC53141 is used to produce scytophycins A, B, C, D and E, which are potent cytotoxins and antineoplastic agents, USA Patent Number5281533
- 84. Schwartz, R.E., Hirsch, C.F., Sesin, D.F., Flor, J.E., Chartrain, M. et al. (1990). Pharmeaceuticals from cultured algae. J Ind Microbiol. 5, 13-123
- 85. Stevenson, C.S., Capper, E.A., Roshak, A.K., Marquez, B., Echman, C. et al. (2002). The identification and characterization of the marine natural product scytonemin as a novel antiproliferative pharmacophore. J Pharmacol Exp Ther. 303, 858-866
- 86. Spolaore, P., Joannis-Cassan, C., Duran, E. and Isambert, A. (2006). Commercial applications of microalgae. J Biosci Bioengrg. 101, 87-96.
- 87. Cardozo, K.H.M., Guaratini, T., Barros, M.O., Falcao, V.R., Tomon, A.P. et al. (2007). Metabolites form algae with economically impact. Comp Biochem Physiol Part C .146, 60-78.
- 88. Stolz, P. and Obermayer, B. (2005). Manufacturing microalgae for skin care. Cosmetics Toiletries, 120,99–106.
- 89. Hillol, C., Shrikrishna, D. J., Dolly W. D. and Sunil, P. (2012). Potential applications of blue green algae. Journal of Scientific & Industrial Research. 71, 13-20
- 90. 90. Priyadarshani, I. and Biswajit, R. (2012). Commercial and industrial applications of microalgae A review J. Algal Biomass Utln. 3, 89–100
- 91. Pulz, O. and Gross, W. (2004). "Valuable products from biotechnology of microalgae." Appl. Microbiol.Biotechnol. 65, 635-648
- 92. Borowitzka, M. A. and Borowitzka, L. J. (1987).Vitamins and Fine Chemicals from Micro-Algae. In M. A. Borowitzka and L. J. Borowitzka (Eds) Micro-Algal Biotechnology, New York, Cambridge University Press.
- 93. Garcia-Gonzalez, M., Moreno, J., Manzano, J.C., Florenzio, F.J., Guererro, M.G. (2005). Production of Dunaliella salina biomass rich in 9-cis-b-carotene and lutein in a closed tubular photobioreactor. J Biotechnol 115, 81-90.
- 94. Leon, R., Martin, M., Vigara, J., Vilchez, C. and Vega, J.M. (2003). Microalgae mediated photoproduction of b-carotene in aqueous-organic two phase systems. Biomol Engrg. 20, 177-183
- 95. Hu, C., Lin, J., Lu, F., Chou, F. and Yang, D.(2008). Determination of carotenoids in Dunaliella salina cultivated in Taiwan and antioxidant capacity of the algal carotenoid extract. Food Chem. 109, 439–446.



- 96. Boussiba, S., Bing, W., Yuan, J.P., Zarka, A., Chen, F. (1999). Changes in pigment profile in the green alga Haematococcus pluvialis exposed to environmental stresses. Biotechnol Lett. 20, 601-604.
- 97. Hejazi, M. and Wijffels, R.H. (2004). Milking microalgae. Trends Biotechnol. 22, 184-194.
- 98. Lorenz, R.T. and Cysewski, G.R. (2000). Commercial potential for Haematococcus microalgae as a natural source of astaxanthin. Trends Biotechnol. 18, 160-167.
- 99. Higuera-Ciapara, I., Felix-Valenzuela, L. and Goycoolea, F.M. (2006). Astaxanthin: a review of its chemistry and applications. Crit Rev Food Sci Nutr. 46, 185-196.
- 100. Yuan, J.P., Peng, J., Yin, K. and Wang, J.H. (2011). Potential health-promoting effects of astaxanthin: a high-value carotenoid mostly from microalgae. Mol Nutr Food Res. 55, 150-165
- 101. Chu, W.L., Phang, S.M., Miyakawa, K. and Tosu, T. (2002). Influence of irradiance and inoculum density on the pigmentation of Spirulina platensis. Asia Pac J Mol Biol Biotechnol. 10, 109–117.
- 102. Borowitzka, M. A. and Borowitzka, L. J. (1987). Vitamins and Fine Chemicals from Micro-Algae. In
- M. A. Borowitzka and L. J. Borowitzka (Eds) Micro- Algal Biotechnology, New York, CambridgeUniversity Press.
- 103. Spolaore, P., Joannis-Cassan, C., Duran, E. and Isambert, A. (2006). Commercial applications of microalgae. J Biosci Bioengrg. 101, 87-96.
- 104. Abe, K., Nishmura, N. and Hirano, M. (1999). Simultaneous production of β-carotene, vitamin E and vitamin C by the aerial microalga Trentepohia aurea. J. Appl. Phycol. 1, 33–6.
- 105. El- Baz, F.K., Aboul-Enein, M.A., El-Baroty, G.S., Youssef, A.M. and Abd El-Baky, H.H. (2002). Accumulation of antioxidant vitamins in Dunaliella salina. Online J. Biol. Sci. 2, 220–223.
 - 106. El-Baky, A.B.D., Moawd, H.H., El-Behairy, A. and ElBaroty, G.S. (2002). Chemoprevention of benzo [a]pyreneinduced carcinogen and lipid peroxidation in mice by lipophilic algae extracts (phycotene). J. Med. Sci. 2, 185–93.
 - 107. Li, Y., Horsman, M., Wu, N., Lan, C.Q. and Dubois-Calero, N. (2008). Biofuels from Microalgae. Biotechnol. Prog. 24, 815-820.
 - 108. Tseng, C. K. (2004). "The past, present and future of phycology in China." Hydrobiologia 512, 11-20.
 - 109. Edwards, M. (2008). Green Algae Strategy End Biowar I and Engineer Sustainable Food and Biofuels. Tempe, Arizona, USA, LuLu Press
 - 110. Lee, Y. K. (1997). "Commercial production of microalgae in the Asia-Pacific rim." J. Appl.Phycol. 9, 403-411
 - 111. Spolaore, P., Joannis-Cassan, C., Duran, E. and Isambert, A. (2006). Commercial applications of microalgae. J. Biosci. Bioeng. 101, 87-96
 - 112. Colla, L.M., Reinehr, C.O., Reichert, C. and Costa, J.A.V. (2007). Production of biomass and nutraceutical compounds by Spirulina platensis under different temperature and nitrogen regimes. Bioresour. Technol. 98, 1489–1493.
 - 113. Sajilata, M.G., Singhal, R.S. and Kamat, M.Y. (2008). Fractionation of lipids and purification of ãlinolenicacid (GLA) from Spirulina platensis. Food Chem. 109, 580–586.
 - 114. Rangel-Yagui, C.O., Danesi, E.D.G., Carvalho, J.C.M. and Sato, S. (2004). Chlorophyll production from Spirulina platensis: cultivation with urea addition by fed-batch process. Bioresour. Technol. 92, 133–141.
 - 115. Madhyastha, H.K. and Vatsala, T.M. 2007 Pigment production in Spirulina fussiformis in different photophysical conditions. Biomol. Eng. 24, 301–305.
 - 116. 116. Ogbonda, K.H., Aminigo, R.E. and Abu, G.O. (2007). Influence of temperature and pH on biomass production and protein biosynthesis in a putative Spirulina sp. Bioresour. Technol. 98, 2207–2211.



- 117. Hills, C. and Nakamura, H. (2006). Food FromSunlight. World Hunger Research Publ., Boulder Creek, CA
- 118. Becker, W. (2004). Microalgae in human and animal nutrition. In A. Richmond (Ed) Handbook of Microalgal Culture, Blackwell, Oxford, pp. 312-351.
- 119. Um, B.H. and Kim, Y.S. (2009). Review: A chance for Korea to advance algal-biodiesel technology. J. Indus. Eng. Chem. 15, 1-7.
- 120. Hallmann, A. (2007). Algal transgenics and biotechnology. Transgenic Plant Journal. 1, 81-98
- 121. Song, T., Martensson, L., Eriksson, T., Zheng, W. and Rasmussen, U. (2005). Biodiversity and seasonal variation of the cyanobacterial assemblage in a rice paddy field in Fujian, China. The Federation of European Materials Societies Microbiology Ecol. 54, 131-140
- 122. Malik, F.R., Ahmed, S. and Rizki Y.M. (2001). Utilization of lignocellulosic waste for the preparation of nitrogenous biofertilizer. Pakistan J. Biol. Sci. 4, 1217–1220
- 123. Priyadarshani, I. and Biswajit, R. (2012). Commercial and industrial applications of microalgae A review. J Algal Biomass Utln. 3, 89–100
- 124. Okuda, K. (2002). "Structure and phylogeny of cellcoverings." J. Plant Res. 115, 283-288.
- 125. Ververis, C., Georghiou K. (2007). "Cellulose, hemicelluloses, lignin and ash content of someorganic materials and their suitability for use as paper pulp supplements." Bioresour. Technol. 98, 296-30.
- 126. Chen, Y.C. (2003). Immobilized Isochrysis galbana (Haptophyta) for long-term storage and applications for feed and water quality control in clam (Meretrix lusoria) cultures. J. Appl. Phycol. 15, 439-444.
- 127. Da Silva, R. L. and Barbosa, J. M. (2008). Seaweed meal as a protein source for the white shrimp Lipopenaeus vannamei. J. Appl. Phycol. 21, 193-197
- 128. http://www.popularmechanics.com/science/energy/a4677/4333722/
- 129. http://algenol.com/direct-to-ethanol/direct-to- ethanol
- 130. http://algenol.com/direct-to-ethanol/environmental-benefits
- 131. "Algenol to change SW Fla., the world".
- 132. "Impending verdict on Algenol's biofuel production utility". Biofuels International, February 2, 2010.
- 133. "Algenol Partners With Lee County as Commission Votes to Approve Incentive Funding for Florida-Based". PR-CANADA.net.
- 134. http://www.solixalgredients.com/
- 135. http://www.sapphireenergy.com/locations/green-crude-farm.html
- 136. FoodNavigator-USA.com. "Solazyme rebrands as TerraVia, raises \$28m as part of plan to focus on food, nutrition, personal care: 'We're redefining the future of food'". FoodNavigator-USA.com. Retrieved 2017-01-24
- 137. "Solazyme Reports Fourth Quarter and Full Year 2014 Results NASDAQ:SZYM)".investors.terravia.com.Retrieved 2017-01-24.
- 138. "Increasing the availability of marine omega-3 -BioMar". www.biomar.com. Retrieved 2017-01-24
- 139. Solazyme (2007-01-11). "Solazyme AnnouncesNew Chief of Genetics". www.prnewswire.com.
- 140. http://en.openei.org/wiki/Aurora_BioFuels_Inc
- 141. http://www.abc.net.au/site-archive/rural/news/content/201211/s3630997.htm
- 142. http://shop.earthrise.com



- 143. http://www.nutrex-hawaii.com/hawaiian-spirulina-pacifica
- 144. www.bulksupplements.com/spirulina-powder.htm
- 145. http://www.taiwanchlorella.com/
- 146. http://www.febico.com
- 147. http://www.nutress.eu/
- 148. http://www.dutchfoodinnovations.com/innovation/essentials-food-natural-algae-rich-food
- 149. http://www.sourcenaturals.com/library/about/
- 150. http://www.herbalhills.in/
- 151. http://www.spirulinaindia.com/
- 152. http://www.parrynutraceuticals.com/
- 153. http://www.pyfarms.com/